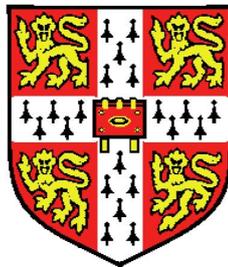


Inference and classification of eukaryotic *cis*-regulatory motifs



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With ♥ to Kaisa.

This dissertation is the result of my own work and includes nothing which is the outcome of work done in collaboration except where specifically indicated in the text.

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Abstract

Regulation of gene expression by networks of sequence specific transcription factors is one of the most important control mechanisms that defines the expression pattern of a genome. Describing transcriptional regulatory networks requires a near complete knowledge of the transcription factors present in the cell, as well the DNA binding sites to which each of the TFs is able to bind. Recent years have witnessed advances in both directions. High coverage transcription factor annotations have become available for many sequenced eukaryotic genomes. Improvements have also been made in profiling DNA specificity motifs for eukaryotic transcription factors, *in vitro* and *in vivo*.

The theme of my work has been the application and development of computational methods for inferring regulatory motifs from promoter sequence, and finding clues to the function of computationally inferred DNA motifs. Functional annotation of inferred motifs led me to conduct a comparative study of the familial relationships between regulatory motifs, the conclusion of which was a probabilistic motif family model I call the ‘metamotif’. The metamotif, I will show, allows improved prediction of the DNA binding domain family for *de novo* inferred motifs, and is an effective way of encoding prior information about known DNA binding domain families to a motif inference algorithm. The use of familial prior information improves the sensitivity to detect regulatory motifs contained in the large promoter sequences that are common to higher eukaryotic genomes. The metamotif guides motif inference towards types of sequence signal that are expected *a priori* to be present in the sequence set of interest, thereby improving and supplementing traditional regulatory motif inference algorithms.

I have also assessed several published *de novo* DNA motif inference algorithms by challenging them to infer a complete set of regulatory motifs from a large series of *Saccharomyces cerevisiae* promoters. This work provides a novel way to assess performance of regulatory motif inference methods, and is made possible by the availability of an experimentally determined regulatory motif dictionary for the *S. cerevisiae* genome. In addition to benchmarking motif inference methods compared to a reference motif set, I make use of many of the rich genomics resources available for study of the budding yeast. These include curated lists of TF target genes based on ChIP-chip and gene expression studies of wild type and knockout yeasts, a close-to-complete list of TF motif from the JASPAR database, and a 7-way sequence conservation score across the genome, as well as sequence variation data from the *Saccharomyces* Genome Resequencing Project.

Development of sensitive regulatory motif inference algorithms continues to be important in gaining understanding of eukaryotic gene regulation by sequence specific transcription factors. In particular I believe that methods that integrate different sources of biological evidence, such as metamotifs, gene expression and ChIP-seq, to sequence motif inference will be highly important to the field.

Contents

Contents	VI
List of Figures	X
1 Introduction	1
1.1 Gene regulation by control of transcription	2
1.1.1 Sequence specific transcription factors	6
1.1.2 Binding specificity of transcription factors	11
1.2 Computational inference of transcription factor binding site motifs	13
1.2.1 The position weight matrix	16
1.3 Computational methodology	18
1.3.1 Hidden Markov Models in motif inference	18
1.3.2 Nested sampling	23
1.3.3 The NestedMICA algorithm	26
1.3.4 Random forest classification	28
1.4 Biological datasets and resources	31
1.4.1 Ensembl	31
1.4.2 Regulatory motif databases	32
1.4.2.1 TRANSFAC	33
1.4.2.2 JASPAR	34
1.4.2.3 UniPROBE	35
1.5 Contributions of this thesis	37
2 Metamotifs - a generative model for building families of nu- cleotide position weight matrices	38

2.1	Background	38
2.1.1	Previous work on motif family models	39
2.2	The metamotif	42
2.2.1	Formulation of the model	45
2.2.2	Visual representation of the model	46
2.2.3	Aligning motifs and estimating metamotifs from a motif multiple alignments	46
2.2.4	Metamotif inference by nested sampling	50
2.2.5	The likelihood function	53
2.2.6	Monte Carlo sampling moves	55
2.2.7	Accounting for incomplete metamotif hits	56
2.3	Evaluating the metamotif nested sampler algorithm	56
2.3.1	A single metamotif	60
2.3.2	Multiple metamotifs	62
2.3.3	Inferring metamotifs from TRANSFAC	63
2.4	Summary	65
3	Metamotifs in motif inference	66
3.1	Previous work on biologically informative motif prior functions . .	67
3.2	Materials & Method	69
3.2.1	The metamotif prior function	69
3.2.2	Measuring motif inference sensitivity with synthetic sequence	71
3.3	Results & Discussion	74
3.3.1	Performance effect of a correct motif family prior function	74
3.3.2	Performance effect of an incorrect motif family prior function	74
3.3.3	Making the metamotif prior available	77
3.3.4	Using the metamotif prior with the NestedMICA algorithm	77
3.3.5	Using the metamotif prior with iMotifs	78
4	Metamotifs in motif classification	81
4.1	Previous work on motif family classification	81
4.2	Materials & Method	83
4.2.1	Training data	84

4.2.2	The classifier feature set	84
4.3	Results & Discussion	85
4.3.1	Performance comparison with previous methods	85
4.3.1.1	MotifPrototyper	86
4.3.1.2	Sparse Multinomial Logistic Regression	88
4.3.2	Performance measurement of two large homeodomain datasets	91
4.3.2.1	Classifying homeodomain motifs by their specificity group	92
4.3.2.2	Clustering of motifs prior to metamotif training .	95
4.3.3	Comparing a metamotif density based classification to a Cartesian distance based classifier	95
4.3.4	Making metamatti available	96
4.3.4.1	The metamatti R package	96
4.3.5	The metamatti web server	97
5	Genome scale motif inference in <i>Saccharomyces cerevisiae</i>	99
5.1	Background	99
5.1.1	Genome scale motif inference	100
5.1.2	Performance inference method assessments	102
5.1.3	The Tompa <i>et al.</i> (2005) assessment	103
5.2	Materials & Method	108
5.2.1	Sequence and annotation retrieval	108
5.2.2	Motif inference	111
5.2.2.1	Unsuccessfully run algorithms	112
5.2.3	Motif comparison	114
5.2.3.1	Motif clustering with the SSD metric	116
5.2.4	Motif scanning	116
5.2.5	Predicted binding site overlap	117
5.2.6	Association of motif hits to transcription factor target genes	118
5.2.6.1	YEASTRACT	119
5.2.6.2	Reimand <i>et al.</i> (2010) TF knockout and expression data based target set	120
5.2.6.3	Harbison <i>et al.</i> (2004) ChIP-chip dataset	120

5.2.6.4	Relationship between discovered motifs and inter-species sequence conservation	121
5.2.7	Relationship between discovered motifs and sequence variation in <i>cerevisiae</i> strains	121
5.2.7.1	Positional bias of motifs	122
5.2.8	Classification of motifs with metamatti	123
5.3	Results & Discussion	124
5.3.1	Properties of inferred motifs	124
5.3.2	Finding matches to known regulatory motifs amongst <i>de novo</i> motif discoveries	128
5.3.3	TF target gene associations of the discovered motifs	141
5.3.4	Clustering of motifs and their binding sites	145
5.3.5	Comparing motifs by the overlap of their genomic matches	149
5.3.6	Looking for evidence of function for the inferred motifs	155
5.3.6.1	Inter-species conservation of the inferred motifs	158
5.3.6.2	SNP rates of the inferred motifs	161
5.3.6.3	Positional bias of motif matches close to the TSS	161
5.3.6.4	Combining the conservation, SNP rate and positional bias to highlight potentially functional motifs	164
5.3.6.5	Classification of the inferred motifs with metamatti	168
5.4	Summary	171
6	Conclusions	173
6.1	Future work	176
Appendix A - iMotifs		182
Appendix B - The motif inference tutorial		188
Appendix C - Motif inference algorithm assessment parameters		196
References		202

List of Figures

1.1	Key regulatory interactions which modulate transcription initiation.	4
1.2	TF counts versus gene counts.	7
1.3	The TF domain coverage of genomes.	10
1.4	Strongly constrained PWM motif, and one with degenerate positions in the middle.	15
1.5	The zero-or-one occurrences per sequence–motif model (ZOOOPS).	21
1.6	The multiple-uncounted sequence-motif mixture model (MUSMM).	22
1.7	The likelihood contour.	24
1.8	The NestedMICA model components: the motif set and the mixing matrix. An ensemble of three states is shown (states labelled 1,2,3).	27
2.1	A forkhead-like metamotif (inferred from an alignment of motifs) is shown alongside selection of motif samples drawn from it.	44
2.2	Visual representations of metamotifs.	47
2.3	Schematic explaining the MLE metamotif inference algorithm.	49
2.4	Example metamotifs for forkhead (A) and HSF (C) motif families from the TRANSFAC database (Matys et al., 2006).	51
2.5	The multiple-uncounted motif–metamotif mixture HMM (MUMM).	54
2.6	Incomplete hits are handled by padding the input motifs with additional columns that fit the background model optimally.	57
2.7	Motifs which were aligned and the multiple alignment summarised as an MLE metamotif with the program <i>nmalign</i> .	59
2.8	Metamotifs estimated with the metamotif nested sampler algorithm with varying relative frequency of metamotif samples.	61

LIST OF FIGURES

2.9	The metamotifs predicted at relative frequency of 0.2 are shown alongside the source metamotifs.	62
2.10	Examples of metamotifs inferred with the nested sampler algorithm from clustered motifs deposited in the TRANSFAC database (Matys et al., 2006).	64
3.1	Metamotif densities with all offsets of the metamotif (shown above the PWM) are summed over the length of the motif.	70
3.2	Synthetic metamotifs contributing to the motif prior functions used in the assessment.	73
3.3	Informative weight matrix prior improves NMICA’s sensitivity to resolve motifs present in human intronic sequence in low frequency (0.2 frequency).	75
3.4	The closest motif match to the invalid motif pattern (ZAP1) shown alongside the ZAP1 motif.	76
3.5	A NestedMICA motif inference run can be configured and run directly in iMotifs.	80
4.1	Accuracy comparison between TF domain superfamily level classification with metamatti and MotifPrototyper (10-fold cross-validation).	87
4.2	Accuracy comparison between the TF domain family classification with metamatti , and SMLR (k-fold cross-validation).	89
4.3	Confusion matrix of the 6-way TRANSFAC motif classification with the metamatti classifier.	90
4.4	Misclassified homeodomain motifs in the A) Noyes et al. (2008a) and the B) Berger et al. (2008) datasets.	92
4.5	Confusion matrix of the homeodomain specificity group classifier. Columns represent the real class, and rows represent the predicted class.	94
4.6	The metamatti motif classification web server.	98
5.1	The sequence retrieval tools included in iMotifs.	110

LIST OF FIGURES

5.2	The number of motifs from different experimental sources in the JASPAR 2010 non-redundant fungal motif dataset.	115
5.3	The ten closest matches between inferred motif sets, and JASPAR motifs. The JASPAR motifs are shown on green background. . . .	126
5.4	Summary of the average lengths and information contents of the different inferred motifs.	128
5.5	The number of statistically significant matches of the predicted motifs with A) JASPAR, and B) Zhu et al. (2009) PBM motifs. . .	129
5.6	The number of reciprocal matches between the predicted motifs and A) JASPAR, and B) Zhu et al. (2009) PBM motifs.	131
5.7	Overlap of significant matches to the JASPAR database between the three top performing motif prediction methods: NestedMICA, MEME and SOMBRERO.	133
5.8	Distribution of SSD distances of predicted motifs to significant matches in the A) JASPAR and B) Zhu et al. (2009) PBM motif sets.	134
5.9	A heatmap showing the JASPAR motifs found or missed by each of the prediction methods ($p < 0.05$).	135
5.10	Different algorithms find matches to partially overlapping subsets of the JASPAR motif set.	137
5.11	JASPAR motifs and computationally predicted motif, grouped according to their A) domain family and B) the motif set.	138
5.12	Differences in length, information content, and column-wise information content between the predicted and the JASPAR reference motifs.	140
5.13	Some <i>de novo</i> inferred motifs are able to distinguish putative TF target genes from non-target genes by the maximum bit scores achieved by the gene promoter sequences (500bp upstream promoter sequences considered).	142
5.14	Motif158 is closely similar to both the CBF1 and PHO4 motifs. . .	143
5.15	TF–target associations of the inferred motifs, when compared to JASPAR motifs.	146

LIST OF FIGURES

5.16	Dendrogram of a complete linkage clustering of all predicted motif sets with the JASPAR motifs, with the SSD metric from Down et al. (2007)	148
5.17	Clustering of JASPAR motifs with results of A) AlignACE, B) Weeder, C) MotifSampler, D) MEME, E) NestedMICA, F) YMF, G)Oligoanalysis H) SOMBRERO.	150
5.18	Numbers of clusters that contain at least one or more inferred, and one or more JASPAR motifs. Four different distance cutoffs are shown.	151
5.19	Motif redundancy as judged by the motif-to-motif SSD distance. A) Fraction of motifs which have at least one pair B) Average motif clique size.	152
5.20	The fraction of motifs with at least one matching pair, at three different significance cutoffs. The consensus string based YMF and Oligoanalysis are omitted from this analysis, because the empirical significance score used here does not behave reliably for PWMs derived from IUPAC consensus strings.	153
5.21	Motif binding site overlap of A) SOMBRERO and B) NestedMICA motifs. The rows represent inferred motifs, and the columns are JASPAR motifs. They are ordered based on an euclidian distance between the overlap patterns, with complete linkage clustering (Johnson, 1967).	154
5.22	Predicted motif similarity to JASPAR motif set on the level of binding site overlap. The bars represent the numbers of motifs which show overlap above 0.10, 0.30, 0.70, 0.90 to JASPAR motifs with the metric described in Section 5.2.5.	155
5.23	The overlap of genomic matches within motif sets. A) SOMBRERO and B) Weeder motifs are shown as examples of the predicted motif sets, and binding site overlap of JASPAR motifs are in panel C. SOMBRERO and Weeder differ in the degree of redundancy amongst the motif set. 500bp upstream sequences were analysed.	156

LIST OF FIGURES

5.24	Predicted motif redundancy on the level of binding site overlap. The bars represent the numbers of motifs which show binding site overlap with the metric described in Section 5.2.5.	157
5.25	Conservation of motifs predicted by NestedMICA.	159
5.26	The number of motifs from each of the predicted motif sets that are found more conserved than intergenic sequence of the same length.	160
5.27	The number of motifs predicted by each of the methods with lower SNP rates than randomly selected intergenic sequence of the matching length. See Section for a description of the bootstrapping based significance scores.	161
5.28	A heat map depiction of the positional bias trends of the motifs inferred with the A) SOMBRERO and B) Weeder algorithms. . .	163
5.29	The fraction of motifs output by each of the eight methods, which show a preference for positions -500 to 0. See Section 5.2.7.1 for details regarding the method.	164
5.30	Overlap of motifs predicted by A) NestedMICA and B) SOMBRERO, that have lower SNP rate than intergenic sequence, higher conservation than intergenic sequence, and are preferential placed within -500 to 0 of TSS.	165
5.31	Motifs predicted by different methods which have lower SNP rate than intergenic sequence, higher conservation than intergenic sequence, and preferential placement close to the TSS.	166
5.32	The ABF1 motif in the JASPAR database. Data originates from the CSI, PBM and Dip-CHIP based study by Badis et al. (2008)	167
5.33	Performance measures of metamatti classification of JASPAR motifs.	169
5.34	Variable importances of a JASPAR family classifier.	170
5.35	Metamatti classification of the predicted motifs at the 0.6 classification probability cutoff.	171
6.1	Three Markov chains aiming to draw a sample from $\mathbb{P}(\mathbf{M} \mathbf{G}, p)$. .	178
6.2	Mixing matrices and their correlations.	179

LIST OF FIGURES

6.3	The sampling algorithm produces mixing matrices that are closely related in correlation pattern to the target (gene expression) correlation matrix.	180
A1	iMotifs can present motif sets and alignments.	184
A2	Output of the nmevaluatebg command plotted in R.	191
A3	The predicted motif alongside known STAT motifs from the TRANS-FAC database.	195
A4	Evaluation of sequence background model class counts at Markov chain order 1.	197
A5	Parameter choices used with Oligo-analysis.	201